Hindawi Journal of Diabetes Research Volume 2018, Article ID 2852398, 6 pages https://doi.org/10.1155/2018/2852398



Review Article

Tubulointerstitial Biomarkers for Diabetic Nephropathy

Bancha Satirapoj 🕞

Division of Nephrology, Department of Medicine, Phramongkutklao Hospital and College of Medicine, Bangkok, Thailand

Correspondence should be addressed to Bancha Satirapoj; satirapoj@yahoo.com

Received 26 October 2017; Revised 29 December 2017; Accepted 4 January 2018; Published 8 February 2018

Academic Editor: Carlos Martinez Salgado

Copyright © 2018 Bancha Satirapoj. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Patients with diabetic nephropathy have a higher risk of mortality, mostly from cardiovascular complications. Standard biomarkers including serum creatinine, estimated glomerular filtration rate, and albuminuria are imprecise, do not directly measure renal tissue injury, and are relatively insensitive to small changes in renal function. Thus, availability of novel biomarkers that are sensitive, specific, and precise as well as able to detect kidney injury and predict clinically significant outcomes would be widely useful in diabetic nephropathy. Novel biomarkers of the processes that induce tubulointerstitial changes may ultimately prove to better predict renal progression and prognosis in type 2 diabetes. Recently, certain biomarkers, which were initially identified in acute kidney injury, also have been reported to confer value in evaluating patients with chronic kidney disease. Biomarkers such as cystatin C, kidney injury molecule-1 (KIM-1), neutrophil gelatinase-associated lipocalin (NGAL), angiotensinogen, periostin, and monocyte chemoattractant protein-1 (MCP-1) reflect tubular injury. In this article, we focused on the potential applications of these biomarkers in diabetic nephropathy.

1. Introduction

In 2010, the worldwide adult population with diabetes mellitus was estimated to be about 285 million and by 2030 an increase by 54% to about 439 million is predicted, reflecting a remarkable increase in diabetes nephropathy [1]. Diabetic nephropathy develops along with generalized microvascular disease, most often concomitant with macrovascular disease including cardiovascular, cerebrovascular, and peripheral arterial diseases [2, 3]. Patients with diabetic nephropathy have a higher risk of mortality, mostly from cardiovascular complications, than diabetic patients without nephropathy [4]. According to the Thailand Renal Replacement Therapy (TRRT) Registry Report 2014, the most common cause of end-stage renal disease (ESRD) is diabetes mellitus.

Diabetic nephropathy is a multifactorial disease involving different pathogenic molecular processes and histopathological structure. A combination of renal tubulointerstitial biomarkers that could capture different pathogenic processes of renal injury may provide a more realistic picture of a patient's outcome. More recently, renal tubulointerstitium has been increasingly reported to play an integral role in the pathogenesis of diabetic nephropathy and correlates well

with progressive renal function decline [5]. Novel biomarkers of the processes that induce tubulointerstitial changes might ultimately prove to be better predictors of renal progression and prognosis in type 2 diabetes (T2DM).

2. Standard Biomarkers of Diabetic Nephropathy

Blood urea nitrogen, serum creatinine, formulas to estimated glomerular filtration rate (GFR), proteinuria, and albuminuria are measures currently used to assess the presence and progress of diabetic nephropathy [2, 3]. However, these measures are imprecise, do not directly measure renal tissue injury, and are relatively insensitive to small changes in renal function. Thus, availability of novel biomarkers that are sensitive, specific, and precise as well as able to detect kidney injury and predict clinically significant outcomes would be widely useful in diabetic nephropathy [6].

Urinary albumin is filtered through the glomerulus and later reabsorbed by tubular cells through the megalincubilin pathway. Albuminuria is an important marker to predict the progression of diabetic nephropathy [7] and more severe and persistent albuminuria correlates with more rapid

progression of diabetic nephropathy and cardiovascular disease among patients with T2DM. Screening for albuminuria should begin at 5-year diabetes duration among patients with type 1 diabetes and at the time of diagnosis among patients with T2DM based on the clinical practice guidelines for diabetic kidney disease outlined by the Kidney Disease Outcomes Quality Initiative (KDOQI) [8]. The preferred screening test is a urine albumin/creatinine ratio with a first-morning void spot collection. A renal biopsy may be necessary to confirm the clinical diagnosis when atypical features are present. Currently, 20-40% of patients with T2DM show GFR that declines before detecting albuminuria [9]. In addition, some studies have shown about 30-45% of diabetic patients with microalbuminuria progressed to macroalbuminuria in 10 years of follow-up; 30% regressed to normoalbuminuria, and 30-40% remained at microalbuminuria level [9]. For these reasons, investigations of new risk markers to predict progression of diabetic nephropathy are suggested.

3. Renal Pathology in Diabetic Nephropathy

Renal pathological changes are present among patients with long-standing diabetes before the onset of microalbuminuria [10]. The characteristic light microscopic features of diabetic nephropathy comprise three major lesions: thickened glomerular basement membranes (GBM) and tubular basement membranes, diffuse mesangial expansion, and hyalinosis of afferent and efferent arterioles. However, hyperglycemiainduced metabolic and hemodynamic stimuli are mediators of kidney injury [11, 12]. These activate inflammatory, prooxidant, ischemic, and fibrotic pathways leading to mesangial matrix accumulation; podocyte effacement and loss; GBM thickening; endothelial dysfunction; tubular atrophy, fibrosis, and dropout; tubulointerstitial inflammation; and renal arteriolar hyalinosis. Therefore, pathology in diabetic nephropathy affects all the renal cellular elements including the glomerular endothelium, mesangial cells, podocytes, and tubular epithelium.

Numerous relationships between tubulointerstitial change and functional outcomes have been reported. Interstitial fibrosis is often proportional to tubular atrophy and a strong predictor of the rate of progression from moderate to severe reduction in GFR [13, 14]. Urinary biomarker data in human beings support the view that tubule injury contributes in a primary way, rather than in a secondary manner, to the development of early diabetic nephropathy [6]. Novel tubular biomarkers related to renal injury in diabetic nephropathy could improve risk stratification and prediction.

4. Markers of Tubulointerstitial Injury

Many of the discovered biomarkers for diabetic nephropathy were identified by transcriptomic and proteomic analyses of renal tissues after injury. As such, a bias exists toward identifying markers from the renal tubulointerstitium, reflecting its greater mass compared with the vascular and glomerular compartments. Recently, certain biomarkers, which were initially identified in acute kidney injury (AKI), also have been reported to confer value in evaluating patients with CKD.

Biomarkers such as cystatin C, kidney injury molecule-1 (KIM-1), neutrophil gelatinase-associated lipocalin (NGAL), angiotensinogen, periostin, and monocyte chemoattractant protein-1 (MCP-1) reflect tubular injury. In this article, we focused on the potential applications of these biomarkers in diabetic nephropathy as shown in Table 1.

4.1. Cystatin-C. Cystatin-C is produced from nucleated cells in the body. It has a molecular weight of 13 kDa, is easily filtered by the glomeruli, and is reabsorbed and catabolized by the proximal tubule. From a prospective observational study, urine cystatin C predicted the progression of T2DM with nephropathy [15]. T2DM with rapid renal progression had significantly increased levels of urine cystatin C when compared with the nonrapid renal progression group [16]. Urine cystatin C was an independent predictor of CKD progression in T2DM.

4.2. Neutrophil Gelatinase-Associated Lipocalin (NGAL). NGAL is a 24 kDa secreted glycoprotein. NGAL is expressed in the renal tubular epithelium, and a rise in urinary concentrations may provide an indication of acute renal injury that can be detected before the rise in serum creatinine concentration [17]. Models of acute kidney injury- (AKI-) to-chronic kidney disease (CKD) transition have implicated its role as a potential biomarker of a chronically injured kidney. In patients with CKD, overexpression of NGAL occurs in renal tubular cells, compared with its expression among non-CKD patients [18], and both serum and urinary NGAL represent a novel, independent renal predictor of the severity of renal disease and CKD progression [19]. In a related study carried out among patients with diabetic nephropathy, elevated urine NGAL level was reported to be associated with the progressive course of the disease leading to ESRD [20]. Similarly, in observational follow-up and our cohort study among patients with T2DM, high urine NGAL levels at baseline correlated with rapid decline of estimated GFR levels and increased serum creatinine [16, 20]. It has been postulated that in the setting of diabetic nephropathy, the rise in NGAL levels occurs independently of decreased GFR and reflects tubular injury and inflammation.

4.3. Kidney Injury Molecule-1 (KIM-1). KIM-1 is a type 1 membrane protein expressed on the apical membrane of proximal tubule cells. Its ectodomain is cleaved and released in the lumen of the tubule and finally appears in urine, which is stable [21]. This biomarker is undetected when the kidneys are normal. Thus, it serves as a specific and sensitive biomarker for proximal tubule damage. In an experimental animal model of tubulointerstitial damage from overload proteinuria, tubular KIM-1 expression was limited to areas with inflammation, fibrosis, and tubular damage [22]. One study also confirmed that KIM-1 was associated with tubulointerstitial inflammation and was overexpressed in the tubules of patients with proteinuric nephropathy, including diabetic nephropathy [23]. From a cross-sectional descriptive study, urine KIM-1 increased in T2DM with normoalbuminuria and mildly increased albuminuria [24]. Serum and urine KIM-1 predicted the rapid decline of GFR [16, 25].

TABLE 1: Biomarkers for diabetic nephropathy.

Biomarkers	Source cell	Description
Standard biomarkers		
Creatinine	Muscle cells	Glomerular filtration markers [8]
		Factors affecting creatinine generation
		(i) extremes of muscle mass,
		(ii) extremes of body size,
		(iii) diet and nutritional status: high protein diet and creatine supplements,
		(iv) muscle wasting diseases.
Albuminuria	_	Glomerular damage markers [8]
		20-40% of diabetic patients with renal impairment exhibited
		normal albuminuria [9]
		No detection in tubulointerstitial injury [9]
Potential biomarkers		
Cystatin C	Nucleated cells/proximal tubular cells	Serum cystatin: glomerular filtration markers
		Urine cystatin: tubular markers
		Predicted the renal progression of type 2 diabetes [15, 16]
Neutrophil gelatinase-associated lipocalin (NGAL)	Neutrophils/distal tubular cells	Urine NGAL: distal tubular markers
		Increased in response to tubulointerstitial injury [17, 18]
		Predicted the renal progression of type 2 diabetes [16, 20]
Kidney injury molecule-1 (KIM-1)	Proximal tubular cells	Urine KIM-1: proximal tubular markers
		Increased in response to tubulointerstitial injury [22, 23]
		Predicted the renal progression of type 2 diabetes [16, 24–26]
Angiotensinogen	Proximal tubular cells	Urine angiotensinogen: proximal tubular markers
		Increased in response to renal RAAS activation [27, 29, 30]
		Predicted the renal progression of type 2 diabetes [16, 28, 31]
Periostin	Bone/distal tubular cells	Urine periostin: distal tubular markers
		Increased in response to renal fibrosis and inflammation [33, 34]
		Predicted the renal progression of type 2 diabetes [36]
Monocyte chemoattractant protein-1 (MCP-1)	Macrophages, glomerular and tubular cells	Urine MCP-1: glomerular and tubular markers
		Increased in response to renal inflammation [39-42]
		Predicted the albuminuria and renal progression of type 2 diabetes [47]

Urinary KIM-1 levels were also significantly higher among patients who progressed from macroalbuminuria to late-stage CKD, but it did not predict progression to end-stage renal disease independently of albuminuria [26]. Urinary KIM-1 excretion could become a noninvasive biomarker of tubulointerstitial injury in diabetic nephropathy. However, Mendelian randomization analysis indicated that the inverse association of increased KIM-1 levels with lower GFR was independent of AER and very likely to represent a causal link [26].

4.4. Angiotensinogen. Angiotensinogen is the only known substrate for renin. Changes in angiotensinogen could influence renin angiotensin aldosterone system (RAAS) activity and increase intrarenal RAAS components, parallel to the severity of fibrotic renal damage, and have been demonstrated in chronic progressive nephropathy [27, 28]. Renal angiotensinogen is formed primarily in proximal tubular cells and is secreted in tubular fluid. The activated intrarenal

RAS was recently proposed to be involved in the progression of renal injury in multiple models of hypertension and in kidney diseases including diabetic nephropathy, immunoglobulin A (IgA) nephropathy, and radiation nephropathy [29, 30]. Urinary angiotensinogen level also correlates with intrarenal angiotensinogen levels and is independently associated with albuminuria and rapid GFR decline in T2DM [16, 28]. In diabetes with normoalbuminuria, urinary angiotensinogen was higher than in controls and demonstrated good performance in differentiating each stage of T2DM from controls [31]. Our research team proposed that angiotensinogen could serve as a potential urinary biomarker to diagnose diabetic nephropathy. It might be useful as an early biomarker of the activation of the RAAS in diabetic nephropathy.

4.5. Periostin. Periostin, osteoblast-specific factor 2, is an extracellular matrix protein firstly expressed in bone, is undetected in other main organs including the kidney [32], and is involved in the fibrosis process and tissue remodeling, kidney

development, and tubular dedifferentiation in experimental models. Our study described, for the first time, the renal expression and urinary excretion of periostin in rats after CKD and in the kidneys of mice with diabetes [33]. Increased periostin expression in the glomeruli and tubular epithelium in diabetic renal pathology was observed. It serves as a marker of renal tubular injury and also correlates with worsening renal outcomes including serum creatinine, blood urea nitrogen, and estimated GFR in various chronic progressive kidney injuries [34, 35]. Urinary periostin levels were more significantly elevated among patients of normoalbuminuria, microalbuminuria, and macroalbuminuria compared with levels of healthy controls. Increased urine periostin level was significantly correlated with aging, high albuminuria, and decline of GFR [36]. The current study indicated that increased urine periostin levels could be detected among patients with T2DM before the onset of significant albuminuria and produced an associated renal derangement among patients with established diabetic nephropathy.

4.6. Monocyte Chemoattractant Protein-1 (MCP-1). The infiltration of inflammatory cells such as monocytes and macrophages in diseased kidneys is a hallmark of the progression of diabetic nephropathy [37]. MCP-1 as a member of the CC chemokine family is a major factor influencing macrophage accumulation in both animal and human models of renal damage [38]. MCP-1 is upregulated and expressed in the diabetic glomerular and renal tubular epithelium [39, 40], and a rise in urinary MCP-1 levels correlates with the extent of interstitial inflammatory infiltrate [41, 42]. In contrast, epidermal growth factor (EGF), a peptide growth factor, plays a protective role in progressive renal injury. EGF plays an important role in restoring barrier function in the healing phase of renal injury and is also a critical in vivo renal cell survival factor for developmentally mature kidneys [43, 44]. Urinary levels of EGF and EGF/MCP-1 ratios are inversely correlated with the extent of tubulointerstitial damage and determined renal prognosis in glomerulonephritis [45, 46]. Urinary levels of MCP-1 among patients with overt nephropathy were also more significantly elevated compared with levels among patients with normal albuminuria [47]. Our preliminary data found that urine MCP-1 and urine EGF/ MCP-1 ratios were independently associated with renal progression among T2DM patients.

5. Conclusion

Novel biomarkers of the processes that induce tubulointerstitial changes may ultimately prove to be better predictors of renal progression and prognosis in T2DM. Our research integrated novel tubular biomarkers simultaneously and compared the performance of each tubular biomarker with standard urine albumin. High levels of urine tubular biomarkers such as urine cystatin C, angiotensinogen, KIM-1, and NGAL presented more rapid decline in renal function among Thai patients with T2DM [16]. This suggests that all novel tubular biomarkers would not be simple surrogate indexes of baseline estimated GFR, but markers on their own, predicting nephropathy progression beyond the

information provided by serum creatinine and other conventional risk factors.

Conflicts of Interest

The author declares that no potential conflict of interests exists.

Acknowledgments

The author would like to thank the medical staff at the Division of Nephrology, Phramongkutklao Hospital and College of Medicine. This study was supported by a grant from Phramongkutklao Hospital and College of Medicine and the National Science and Technology Development Agency (NSTDA, P-13-00505), Bangkok, Thailand.

References

- [1] G. Tramonti and Y. S. Kanwar, "Review and discussion of tubular biomarkers in the diagnosis and management of diabetic nephropathy," *Endocrine*, vol. 43, no. 3, pp. 494–503, 2013
- [2] B. Satirapoj and S. G. Adler, "Comprehensive approach to diabetic nephropathy," *Kidney Research and Clinical Practice*, vol. 33, no. 3, pp. 121–131, 2014.
- [3] B. Satirapoj and S. G. Adler, "Prevalence and management of diabetic nephropathy in western countries," *Kidney Diseases*, vol. 1, no. 1, pp. 61–70, 2015.
- [4] M. Afkarian, M. C. Sachs, B. Kestenbaum et al., "Kidney disease and increased mortality risk in type 2 diabetes," *Journal of the American Society of Nephrology*, vol. 24, no. 2, pp. 302–308, 2013.
- [5] C. E. Hills and P. E. Squires, "The role of TGF-β and epithelialto mesenchymal transition in diabetic nephropathy," *Cytokine & Growth Factor Reviews*, vol. 22, no. 3, pp. 131–139, 2011.
- [6] B. Satirapoj, C. C. Nast, and S. G. Adler, "Novel insights into the relationship between glomerular pathology and progressive kidney disease," *Advances in Chronic Kidney Disease*, vol. 19, no. 2, pp. 93–100, 2012.
- [7] H.-H. Parving, B. Oxenboll, P. A. Svendsen, J. S. Christiansen, and A. R. Andersen, "Early detection of patients at risk of developing diabetic nephropathy. A longitudinal study of urinary albumin excretion," *Acta Endocrinol*, vol. 100, no. 4, pp. 550–555, 1982.
- [8] National Kidney Foundation, "KDOQI clinical practice guideline for diabetes and CKD: 2012 update," *American Journal of Kidney Diseases*, vol. 60, no. 5, pp. 850–886, 2012.
- [9] P. Fioretto, M. W. Steffes, and M. Mauer, "Glomerular structure in nonproteinuric IDDM patients with various levels of albuminuria," *Diabetes*, vol. 43, no. 11, pp. 1358–1364, 1994.
- [10] S. G. Adler, S. W. Kang, S. Feld et al., "Can glomerular mRNAs in human type 1 diabetes be used to predict transition from normoalbuminuria to microalbuminuria?," *American Journal of Kidney Diseases*, vol. 40, no. 1, pp. 184–188, 2002.
- [11] B. Satirapoj, "Review on pathophysiology and treatment of diabetic kidney disease," *Journal of the Medical Association of Thailand*, vol. 93, Supplement 6, pp. S228–S241, 2010.
- [12] B. Satirapoj, "Nephropathy in diabetes," *Advances in Experimental Medicine and Biology*, vol. 771, pp. 107–122, 2012.

[13] P. Fioretto, M. W. Steffes, D. E. Sutherland, and M. Mauer, "Sequential renal biopsies in insulin-dependent diabetic patients: structural factors associated with clinical progression," Kidney International, vol. 48, no. 6, pp. 1929–1935, 1995.

- [14] S. Adler, L. J. Striker, G. E. Striker, D. T. Perkinson, J. Hibbert, and W. G. Couser, "Studies of progressive glomerular sclerosis in the rat," *The American Journal of Pathology*, vol. 123, no. 3, pp. 553–562, 1986.
- [15] S. S. Kim, S. H. Song, I. J. Kim et al., "Urinary cystatin C and tubular proteinuria predict progression of diabetic nephropathy," *Diabetes Care*, vol. 36, no. 3, pp. 656–661, 2013.
- [16] B. Satirapoj, K. Aramsaowapak, T. Tangwonglert, and O. Supasyndh, "Novel tubular biomarkers predict renal progression in type 2 diabetes mellitus: a prospective cohort study," *Journal of Diabetes Research*, vol. 2016, Article ID 3102962, 9 pages, 2016.
- [17] J. Mishra, K. Mori, Q. Ma et al., "Amelioration of ischemic acute renal injury by neutrophil gelatinase-associated lipocalin," *Journal of the American Society of Nephrology*, vol. 15, no. 12, pp. 3073–3082, 2004.
- [18] A. Viau, K. El Karoui, D. Laouari et al., "Lipocalin 2 is essential for chronic kidney disease progression in mice and humans," *Journal of Clinical Investigation*, vol. 120, no. 11, pp. 4065– 4076, 2010.
- [19] D. Bolignano, A. Lacquaniti, G. Coppolino et al., "Neutrophil gelatinase-associated lipocalin (NGAL) and progression of chronic kidney disease," *Clinical Journal of the American Society of Nephrology*, vol. 4, no. 2, pp. 337–344, 2009.
- [20] Y. H. Yang, X. J. He, S. R. Chen, L. Wang, E. M. Li, and L. Y. Xu, "Changes of serum and urine neutrophil gelatinase-associated lipocalin in type-2 diabetic patients with nephropathy: one year observational follow-up study," *Endocrine*, vol. 36, no. 1, pp. 45–51, 2009.
- [21] H. J. Bangstad, I. Seljeflot, T. J. Berg, and K. F. Hanssen, "Renal tubulointerstitial expansion is associated with endothelial dysfunction and inflammation in type 1 diabetes," *Scandinavian Journal of Clinical and Laboratory Investigation*, vol. 69, no. 1, pp. 138–144, 2009.
- [22] M. M. van Timmeren, S. J. Bakker, V. S. Vaidya et al., "Tubular kidney injury molecule-1 in protein-overload nephropathy," *American Journal of Physiology-Renal Physiology*, vol. 291, no. 2, pp. F456–F464, 2006.
- [23] M. M. van Timmeren, M. C. van den Heuvel, V. Bailly, S. J. Bakker, H. van Goor, and C. A. Stegeman, "Tubular kidney injury molecule-1 (KIM-1) in human renal disease," *The Journal of Pathology*, vol. 212, no. 2, pp. 209–217, 2007.
- [24] J. A. de Carvalho, E. Tatsch, B. S. Hausen et al., "Urinary kidney injury molecule-1 and neutrophil gelatinase-associated lipocalin as indicators of tubular damage in normoalbuminuric patients with type 2 diabetes," *Clinical Biochemistry*, vol. 49, no. 3, pp. 232–236, 2016.
- [25] S. E. Nielsen, H. Reinhard, D. Zdunek et al., "Tubular markers are associated with decline in kidney function in proteinuric type 2 diabetic patients," *Diabetes Research and Clinical Prac*tice, vol. 97, no. 1, pp. 71–76, 2012.
- [26] N. M. Panduru, N. Sandholm, C. Forsblom et al., "Kidney injury molecule-1 and the loss of kidney function in diabetic nephropathy: a likely causal link in patients with type 1 diabetes," *Diabetes Care*, vol. 38, no. 6, pp. 1130–1137, 2015.
- [27] M. L. Graciano, C. Cavaglieri Rde, H. Delle et al., "Intrarenal renin-angiotensin system is upregulated in experimental

- model of progressive renal disease induced by chronic inhibition of nitric oxide synthesis," *Journal of the American Society of Nephrology*, vol. 15, no. 7, pp. 1805–1815, 2004.
- [28] S. S. Kim, S. H. Song, I. J. Kim et al., "Clinical implication of urinary tubular markers in the early stage of nephropathy with type 2 diabetic patients," *Diabetes Research and Clinical Practice*, vol. 97, no. 2, pp. 251–257, 2012.
- [29] Y. Suzaki, Y. Ozawa, and H. Kobori, "Intrarenal oxidative stress and augmented angiotensinogen are precedent to renal injury in Zucker diabetic fatty rats," *International Journal of Biological Sciences*, vol. 3, no. 1, pp. 40–46, 2007.
- [30] K. Miyata, N. Ohashi, Y. Suzaki, A. Katsurada, and H. Kobori, "Sequential activation of the reactive oxygen species/angiotensinogen/renin-angiotensin system axis in renal injury of type 2 diabetic rats," *Clinical and Experimental Pharmacology and Physiology*, vol. 35, no. 8, pp. 922–927, 2008.
- [31] B. Satirapoj, N. Siritaweesuk, and O. Supasyndh, "Urinary angiotensinogen as a potential biomarker of diabetic nephropathy," *Clinical Kidney Journal*, vol. 7, no. 4, pp. 354–360, 2014.
- [32] S. Takeshita, R. Kikuno, K. Tezuka, and E. Amann, "Osteoblast-specific factor 2: cloning of a putative bone adhesion protein with homology with the insect protein fasciclin I," *Biochemical Journal*, vol. 294, no. 1, pp. 271–278, 1993.
- [33] B. Satirapoj, Y. Wang, M. P. Chamberlin et al., "Periostin: novel tissue and urinary biomarker of progressive renal injury induces a coordinated mesenchymal phenotype in tubular cells," *Nephrology Dialysis Transplantation*, vol. 27, no. 7, pp. 2702–2711, 2012.
- [34] P. Wantanasiri, B. Satirapoj, M. Charoenpitakchai, and P. Aramwit, "Periostin: a novel tissue biomarker correlates with chronicity index and renal function in lupus nephritis patients," *Lupus*, vol. 24, no. 8, pp. 835–845, 2015.
- [35] B. Satirapoj, R. Witoon, P. Ruangkanchanasetr, P. Wantanasiri, M. Charoenpitakchai, and P. Choovichian, "Urine periostin as a biomarker of renal injury in chronic allograft nephropathy," *Transplantation Proceedings*, vol. 46, no. 1, pp. 135– 140, 2014.
- [36] B. Satirapoj, S. Tassanasorn, M. Charoenpitakchai, and O. Supasyndh, "Periostin as a tissue and urinary biomarker of renal injury in type 2 diabetes mellitus," *PLoS One*, vol. 10, no. 4, article e0124055, 2015.
- [37] T. Furuta, T. Saito, T. Ootaka et al., "The role of macrophages in diabetic glomerulosclerosis," *American Journal of Kidney Diseases*, vol. 21, no. 5, pp. 480–485, 1993.
- [38] S. Segerer, P. J. Nelson, and D. Schlöndorff, "Chemokines, chemokine receptors, and renal disease: from basic science to pathophysiologic and therapeutic studies," *Journal of the American Society of Nephrology*, vol. 11, no. 1, pp. 152–176, 2000.
- [39] T. Wada, K. Furuichi, N. Sakai et al., "Up-regulation of monocyte chemoattractant protein-1 in tubulointerstitial lesions of human diabetic nephropathy," *Kidney International*, vol. 58, no. 4, pp. 1492–1499, 2000.
- [40] N. Banba, T. Nakamura, M. Matsumura, H. Kuroda, Y. Hattori, and K. Kasai, "Possible relationship of monocyte chemoattractant protein-1 with diabetic nephropathy," *Kidney International*, vol. 58, no. 2, pp. 684–690, 2000.
- [41] G. Grandaliano, L. Gesualdo, E. Ranieri et al., "Monocyte chemotactic peptide-1 expression in acute and chronic human nephritides: a pathogenetic role in interstitial monocytes recruitment," *Journal of the American Society of Nephrology*, vol. 7, no. 6, pp. 906–913, 1996.

[42] B. H. Rovin, N. Doe, and L. C. Tan, "Monocyte chemoattractant protein-1 levels in patients with glomerular disease," American Journal of Kidney Diseases, vol. 27, no. 5, pp. 640–646, 1996.

6

- [43] J. Lechner, N. A. Malloth, P. Jennings, D. Heckl, W. Pfaller, and T. Seppi, "Opposing roles of EGF in IFN-α-induced epithelial barrier destabilization and tissue repair," *American Journal of Physiology-Cell Physiology*, vol. 293, no. 6, pp. C1843–C1850, 2007.
- [44] W. A. Kennedy 2nd, R. Buttyan, E. Garcia-Montes, C. A. Olsson, and I. S. Sawczuk, "Epidermal growth factor suppresses renal tubular apoptosis following ureteral obstruction," *Urology*, vol. 49, no. 6, pp. 973–980, 1997.
- [45] S. Worawichawong, S. Worawichawong, P. Radinahamed et al., "Urine epidermal growth factor, monocyte chemoattractant protein-1 or their ratio as biomarkers for interstitial fibrosis and tubular atrophy in primary glomerulonephritis," *Kidney and Blood Pressure Research*, vol. 41, no. 6, pp. 997– 1007, 2016.
- [46] L. Gesualdo, S. Di Paolo, A. Calabró et al., "Expression of epidermal growth factor and its receptor in normal and diseased human kidney: an immunohistochemical and in situ hybridization study," Kidney International, vol. 49, no. 3, pp. 656–665, 1996.
- [47] T. Morii, H. Fujita, T. Narita et al., "Association of monocyte chemoattractant protein-1 with renal tubular damage in diabetic nephropathy," *Journal of Diabetes and its Complications*, vol. 17, no. 1, pp. 11–15, 2003.

















Submit your manuscripts at www.hindawi.com







Computational and Mathematical Methods in Medicine



Behavioural Neurology











